



***Simultaneous removal of aggregates,
leached protein A, endotoxin, and
DNA from protein A purified IgG with
CHT™ ceramic hydroxyapatite and
CFT™ ceramic fluorapatite***

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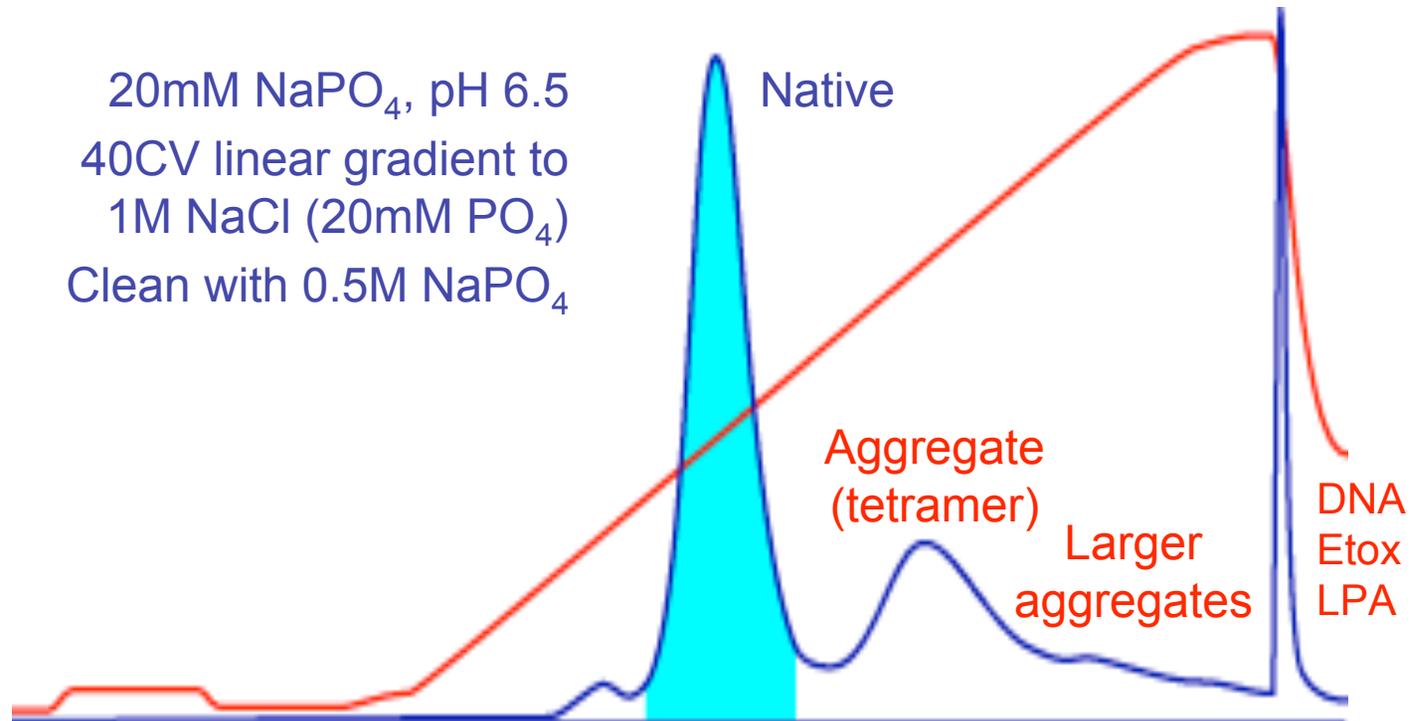
Purification of Biological Products, Santa Monica, December 5–7, 2005



Chemical structure, hydroxyapatite

- **Calcium hydroxyapatite**
- $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$
- Calcium participates in metal affinity interactions
- Phosphate participates in cation exchange/exclusion interactions
- Stable down to pH 6.5 in the presence of 5mM phosphate

CHT fractionation of contaminants



Protein A-purified human IgG1
CHT type I, 20 micron, 300 cm/hr

Analysis of CHT fractions

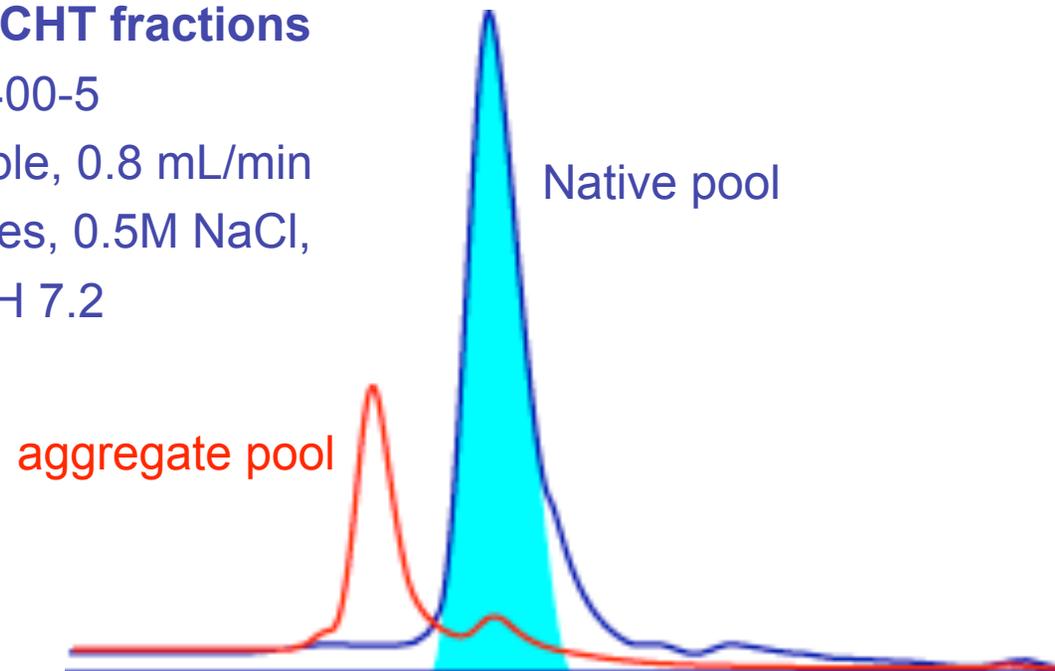
HPSEC of CHT fractions

Bio-Silect 400-5

50 μ L sample, 0.8 mL/min

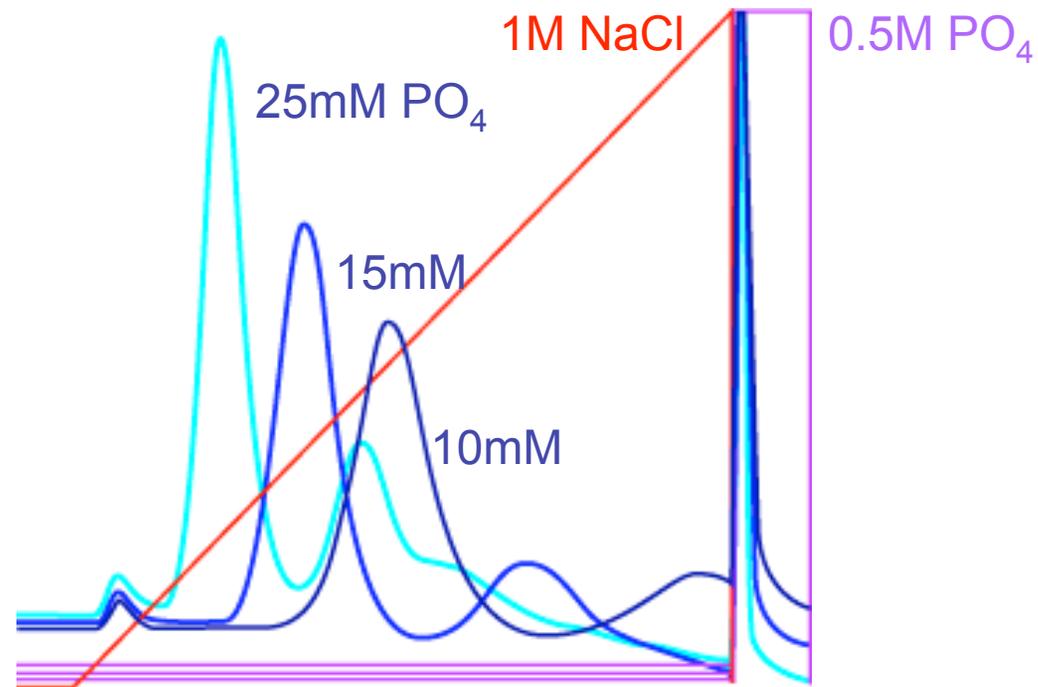
50mM HEPES, 0.5M NaCl,

2M urea, pH 7.2



The effect of PO₄ on CHT

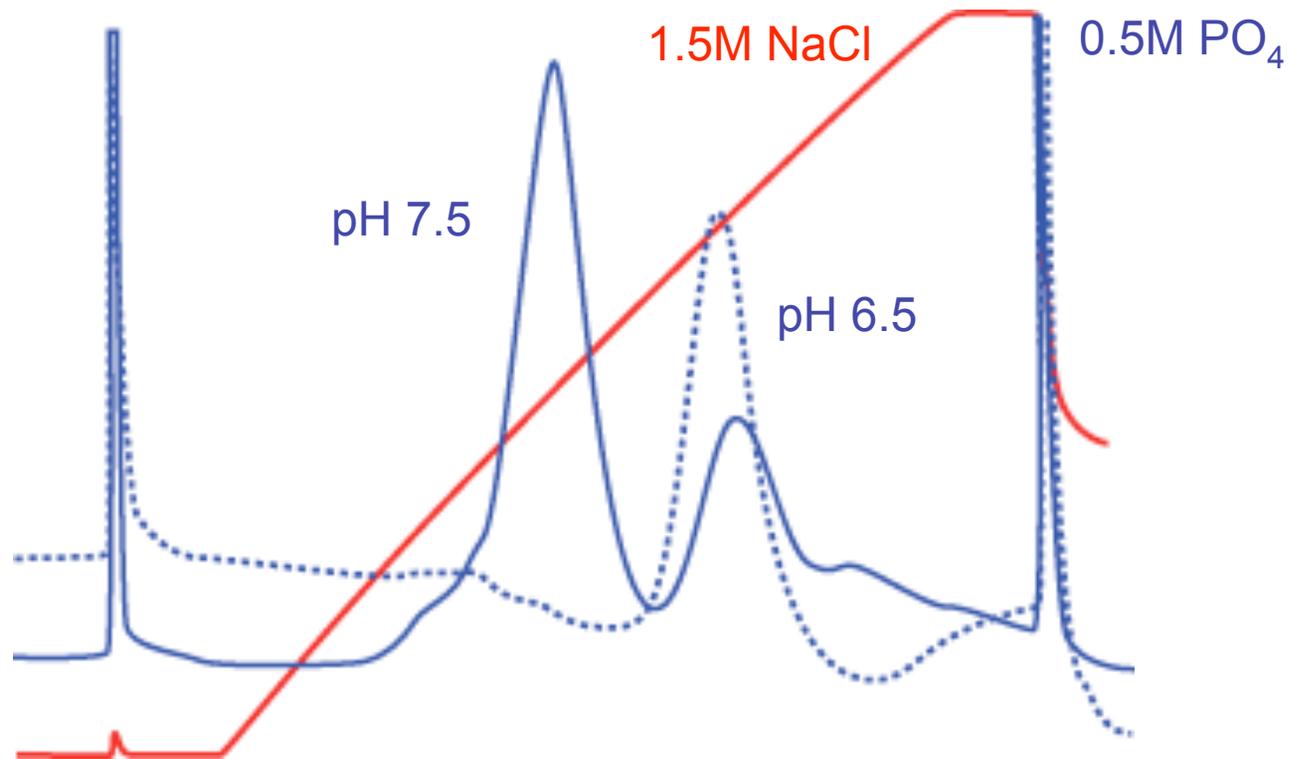
Indicated phosphate concentration maintained across the sodium chloride gradient



protein A purified IgG on CHT type I 20 μm

The effect of pH on CHT

Sodium chloride gradient at constant 5mM NaPO₄



protein A purified IgG on CHT type I 20 µm

Summary of CHT performance

- **Aggregate removal**
 - > 99% by HPSEC
 - from > 40% to < 1%
 - **Leached protein A removal**
 - 90% by Cygnus*
 - from 55 to 5 ng/mL
 - **DNA removal**
 - > 3 logs by PCR
 - down to < 1ng/mL by picogreen
 - **Endotoxin removal**
 - 7×10^4 by LAL
 - down to 1EU/mL
- * at 20 mM NaPO₄, >99% LPA removal at 5mM

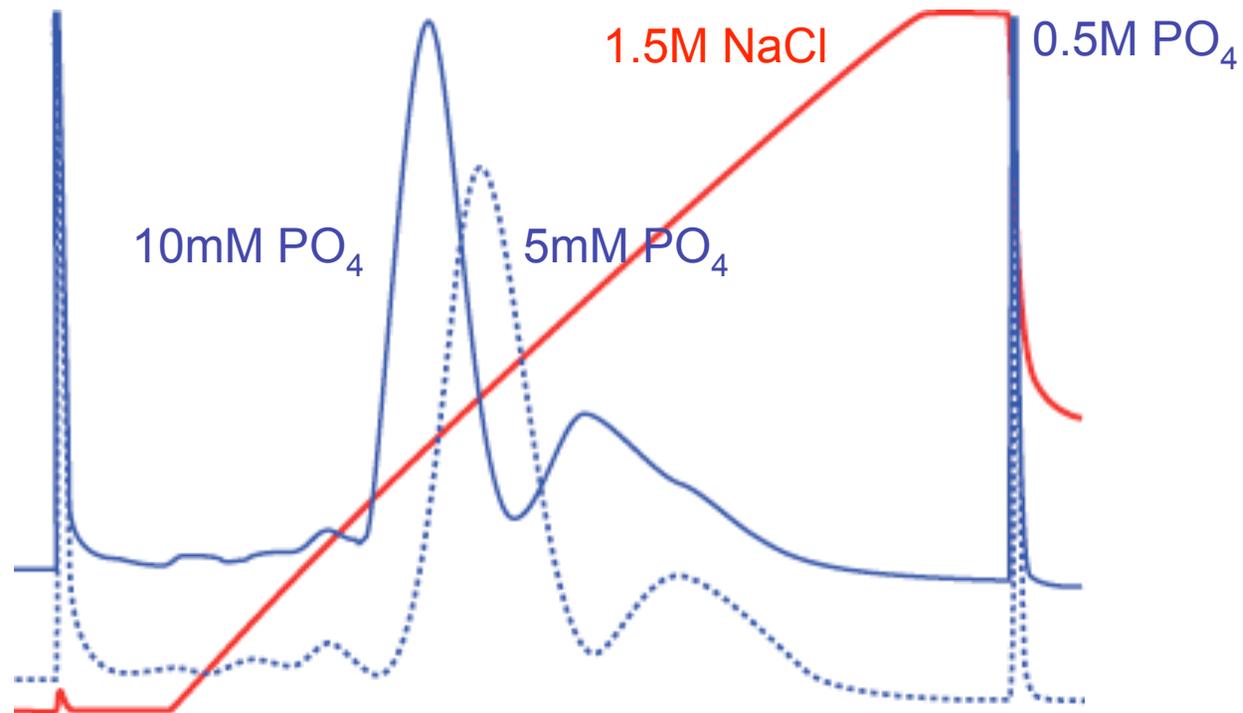


Chemical structure, fluorapatite

- **Calcium fluorapatite**
- $\text{Ca}_{10}(\text{PO}_4)_6\text{F}_2$
- Calcium participates in metal affinity interactions
- Phosphate participates in cation exchange/exclusion interactions
- Stable to pH 5.5
- 4-5 times more mechanically stable than CHT

The effect of PO₄ on CFT

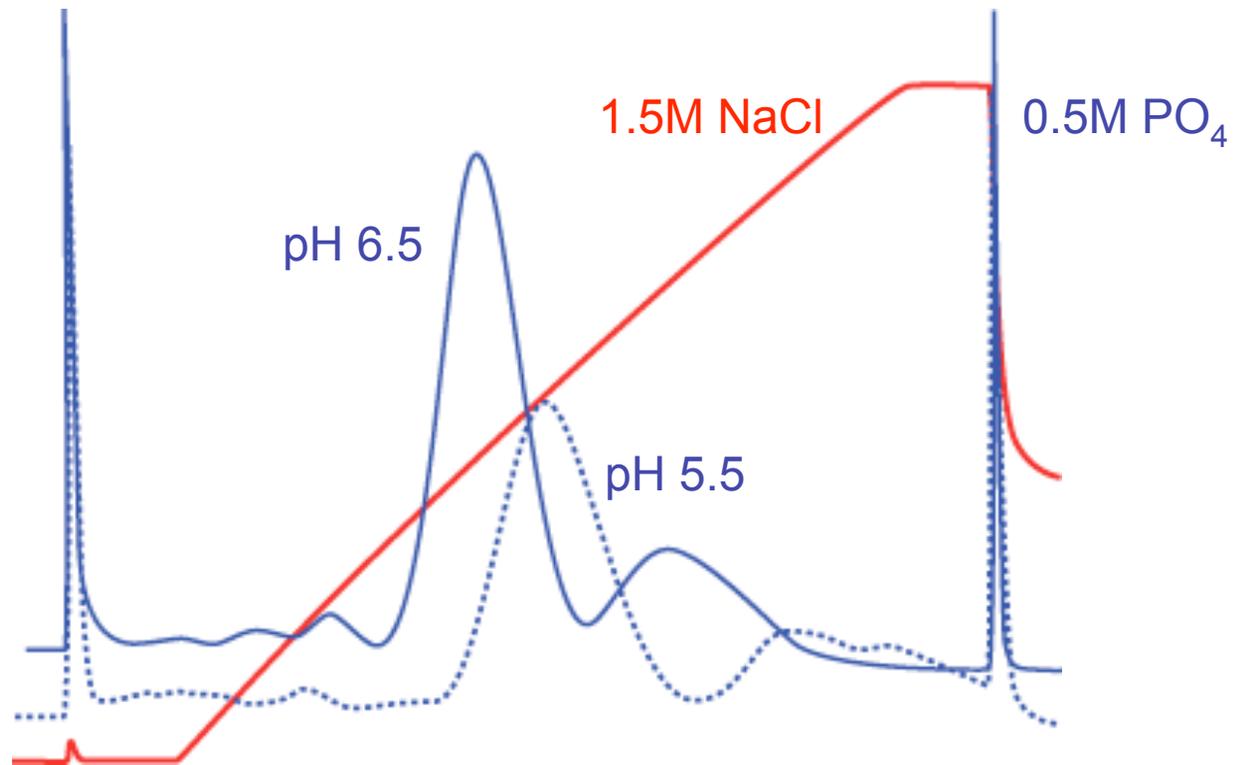
Sodium chloride gradient pH 6.5



protein A purified IgG on CFT type II 40 μm

The effect of pH on CFT

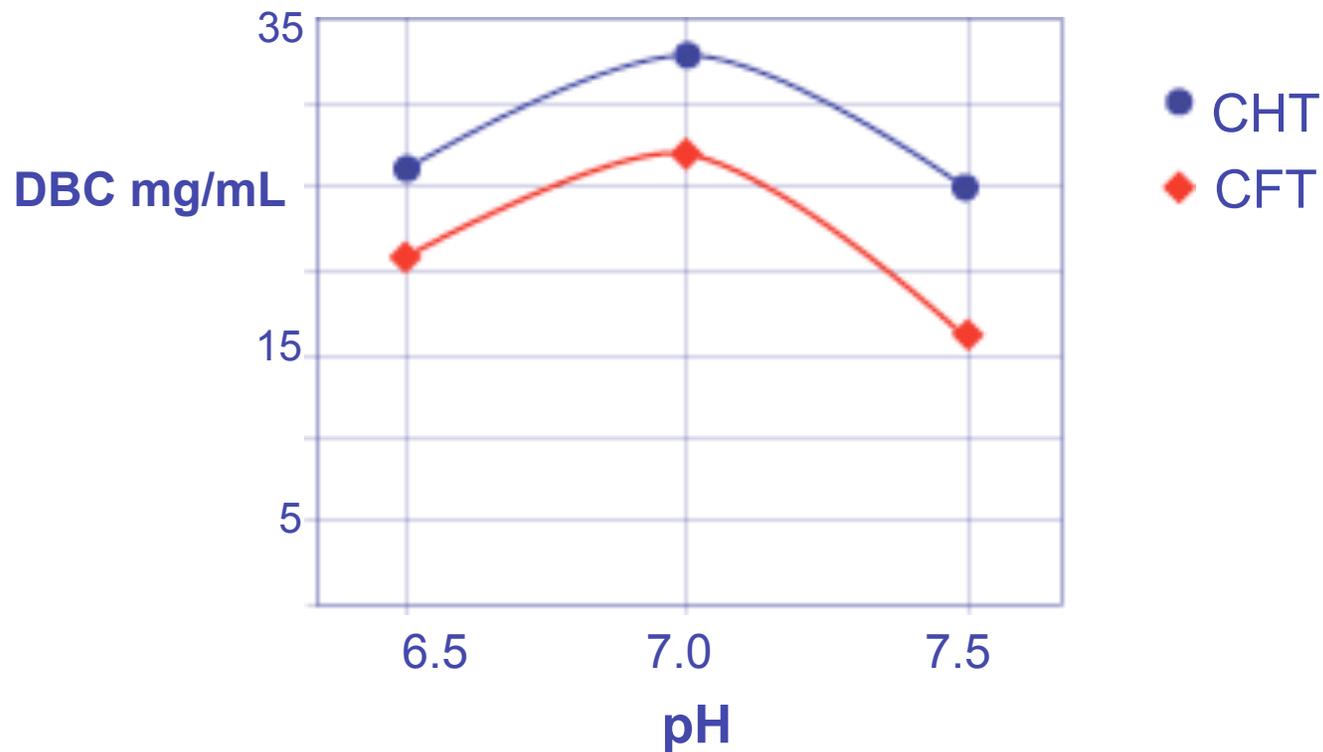
Sodium chloride gradient at 5mM NaPO₄



protein A purified IgG on CFT type II 40 µm

Capacity CHT vs CFT

Dynamic binding capacity of polyclonal human IgG
on CHT type I 40 μm and CFT type II 40 μm
10% breakthrough, 300 cm/hr



Contaminant removal CHT vs CFT

	CHT, type I, 40 μ m			CFT, type II, 40 μ m		
[PO ₄] mM	5	10	15	5	10	15
PA ng/mL	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
DNA ng/mL	<1.0	<1.0	3.9	1.7	<1.0	<1.0
Etox, EU/mL	<0.05	1.0	1.6	3.9	6.2	9.5

Sample: protein A purified IgG. 22 ng/mL leached protein A, 2.3×10^3 ng/mL DNA, 1.9×10^4 EU/mL endotoxin

All results for a sodium chloride gradient to 1.5 M at pH 6.5 with phosphate concentration held at the indicated level, followed by a cleaning step at 0.5 M NaPO₄, pH 6.5

Initial screening conditions CFT/CHT

- Equilibrate column with 5 mM NaPO₄, pH 6.5
- Inject 5% CV protein A purified IgG
- Wash 5 CV equilibration buffer
- Elute 30 CV linear gradient to 1.5 M NaCl (5mM NaPO₄)
- Clean with 0.5M NaPO₄.

- If native MAb peak fails to elute within the NaCl gradient, raise the phosphate concentration enough to bring it in (increments of 5mM or less).
- Optimize by adjusting slope and amplitude of NaCl gradient. Convert to step or flow-through for scale-up.



2-Step platform, protein A/CHT

- Elute protein A with 0.1M glycine* or arginine* 0.05M NaCl, pH 3.8 (no citrate or EDTA).
- Hold for viral inactivation.
- Raise pH to 6.5 by addition of 0.5M NaPO₄ pH 10.5, 1% v:v.
- Equilibrate CHT to 5mM NaPO₄, pH 6.5
- Run optimized CHT fractionation conditions

* Glycine and arginine concentration can be raised to 1-2M to reduce aggregation. Both are dielectric constant enhancers preferentially excluded from protein surfaces. They improve solubility at the same time that they stabilize proteins. Since both are zwitterionic above pH 5 they contribute nothing to conductivity when neutralized.

2-Step platform, protein A/CHT

	OM	PPA	PCHT
Aggregate %IgG	-----	>40	<1
Protein A ng	-----	162	6
DNA ng	9.9x10 ⁵	3.8x10 ⁴	12
Endotoxin EU	2.6x10 ³	5.0x10 ²	<0.05
IgG %	100	25*	45*

OM: original material

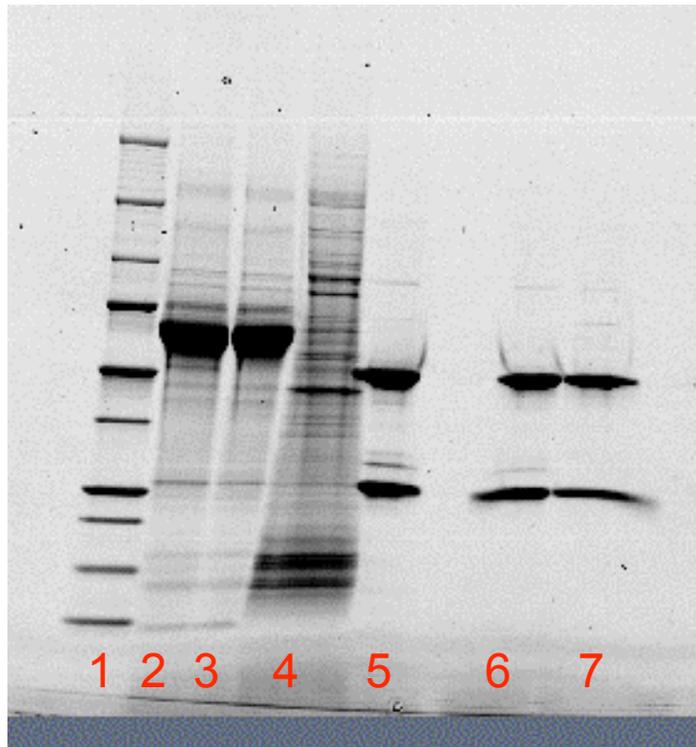
PPA: IgG pool from protein A

PCHT: native IgG pool from CHT

*low recovery PPA due to aggregation; PCHT due to aggregate removal

2-Step platform, protein A/CHT

Reduced SDS PAGE (Flamingo stain*)



1. MW stds
2. OM
3. PA flow-through
4. PA wash (KS)
5. PA pool
6. CHT native pool
7. CHT aggregate pool

KS: 1M NaCl, 2M urea,
10mM EDTA, 0.05M PO₄

*sensitivity equivalent to silver



Conclusions

- CHT and CFT, when eluted with a sodium chloride gradient at a low concentration of phosphate, have a unique ability to simultaneously achieve major reductions in the levels of aggregates, leached protein A, DNA, and endotoxin.
- Scouting/feasibility can often be covered by a single experiment with a sodium chloride gradient at 5mM NaPO₄ pH 6.5; sometimes another with 10mM. Increments of pH can also be investigated.
- The method is easily integrated into a 2-step platform with protein A, or with a variety of 3-step platforms that exploit additional fractionation mechanisms.



Conclusions

- The selectivities of CHT and CFT, although similar and based on the same mechanisms, are distinct.
- CHT supports better resolution from endotoxin but performance is roughly equivalent for removal of aggregates, protein A, and DNA.
- CHT supports about 20% higher capacity
- CFT is more stable chemically and mechanically.

Acknowledgments

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- Thanks also to Rolf Frey, Doug Pagano, Russ Frost, Ursula Snow, Tetsuro Ogawa, and Professor Tsuneo Okuyama for many stimulating discussions.
- For a copy of this presentation, or other Bio-Rad resources concerning CHT or CFT, please contact <andrew_cohen@bio-rad.com>
- For technical questions concerning application development, you are welcome to contact <peter_gagnon@bio-rad.com>